

## Redox-Responsive Controlled Gene Transfection Based on Polymer-Conjugated Magnetic Nanoparticles

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### Abstract

Gene transfection is a non-viral therapy on gene-based diseases by delivering nucleic acids into the nucleus of target cells<sup>1</sup>. The efficiency of gene transfection may be enhanced by magnetofection which involves magnetic nanomaterials (MNPs) under a magnetic field<sup>2</sup>. To combine nucleic acids with nanoparticles as well as protect them from degradation after endocytosis, MNPs are usually modified with cationic compounds, such as 25 kDa branched polyethylenimine (PEI)<sup>3,4</sup>. After cationic adsorption of plasmid DNAs on the surface of negative charged MNPs, addition of extra free PEI is often required to form a ternary complex for magnetofection<sup>5</sup>. It is because only the cationic compounds could transfer the nucleic acids into the cell nucleus, while the MNPs stay only in the perinuclear region. In this work, a redox-responsive disulfide bond is used to link 25 kDa PEI to MNPs, generating detachable PEIs for both DNA protection and nuclear entry.

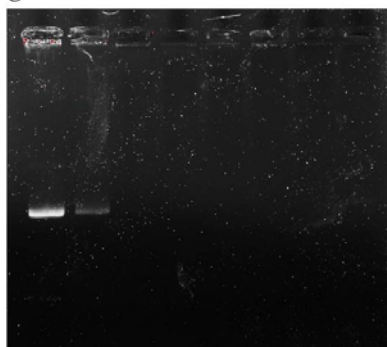
The as-synthesized MNPs were first wrapped in silica with thiol groups on the surface. After thiol-exchanging with 2-carboxyethyl-2-pyridyl disulfide, PEI was linked to the carboxyl groups with EDC/NHS. The results of agarose gel electrophoresis indicated that 10  $\mu\text{g}$   $\text{Fe}_3\text{O}_4$  nanoparticles could condense 250 ng pRL-CMV, *i.e.*, renilla luciferase control reporter vectors (**Fig. 1**). Repeated experiments indicated that the condensing nanoparticles were stable for at least one month. The magnetic gene carrier exhibited efficient gene transfection in both Hela and HepG2 cells lines (**Fig. 2**). After addition of 10 mM glutathione (GSH) in phosphate buffer solution (pH=7.4), plasmid DNA was released from the nanoparticles, confirming the redox-responsive property of the modified magnetic nanoparticles (**Fig. 3**).

The confocal microscopy images showed the labeled plasmid DNA located in the nucleus 3h post-transfection, which was more obvious 24 h after transfection (**Fig. 4**). In addition, the yellow colored area indicated a significant portion of PEI co-localized with the red florescent lysotracker, suggesting that the magnetic nanoparticles were taken into the cells via the endocytosis pathway. The co-localization of PEI and plasmid DNA in the nucleus confirmed the nucleic acids were taken in with the help of PEI, while nanoparticles were still in the perinuclear region.

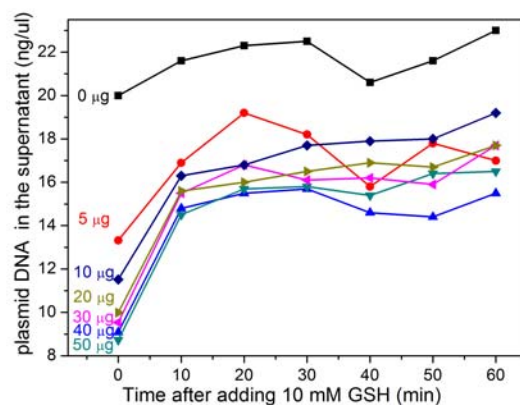
### References

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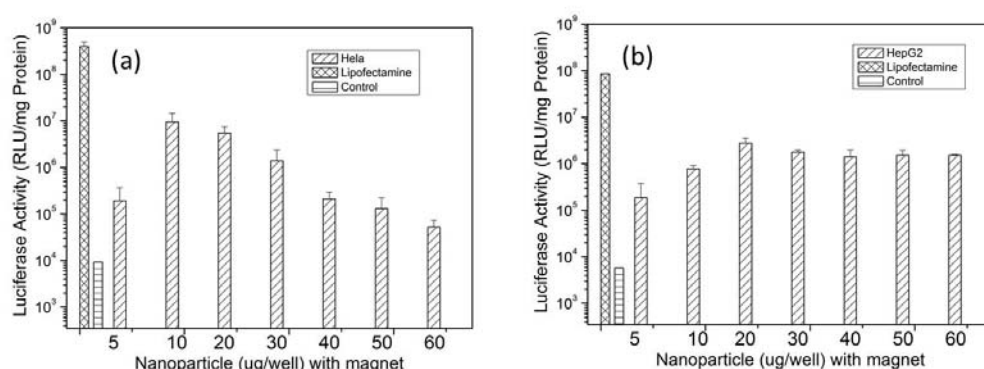
NP/ $\mu\text{g}$  0 5 10 15 20 30 40 50



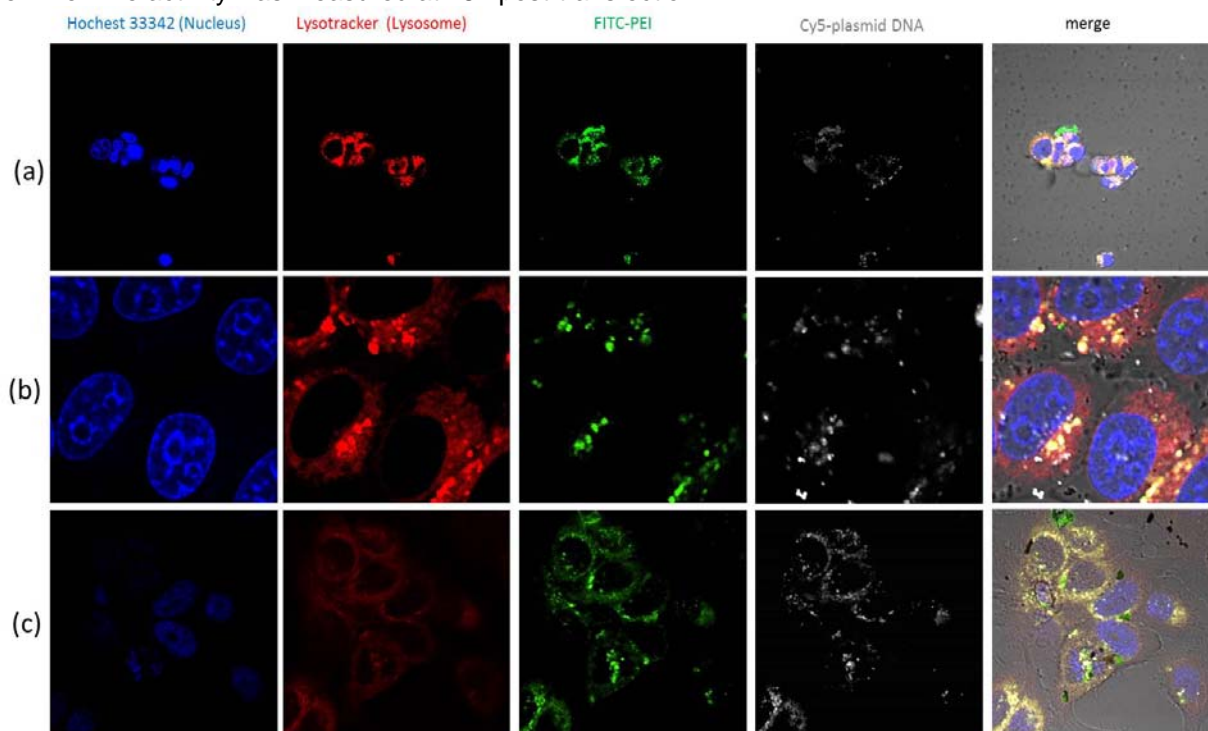
**Fig1.** Evaluation of the DNA condensing ability of  $\text{sFe}_3\text{O}_4@\text{SiO}_2\text{-SS-PEI}$  with agarose gel electrophoresis gel assay. Different weight of nanoparticles was incubated with 250 ng pRL-CMV for 20 min at room temperature.



**Fig 2.** pRL-CMV released at different endpoints after addition of 10mM GSH. Plasmid DNA was mixed with different amount of  $s\text{Fe}_3\text{O}_4@\text{SiO}_2\text{-SS-PEI}$ , and magnet was used to separate the condensed DNA on nanoparticles with the DNA in the supernatant.



**Fig 3.** Evaluation of the transfection efficiency using luciferase assay: (a) in HeLa cell line; (b) in HepG2 cell line. The activity was measured at 48h post-transfection.



**Fig 4.** Tracking of different components in HeLa cells 1 h (a), 3 h (b) and 24 h (c) post-magnetofection. Nucleus was labeled by Hoechst 33342, lysosome with LysoTracker® Red DND-99, PEI with FITC, and plasmid DNA with Cy5. Merged images with bright field were also shown in the right column.